# WG1: Fertility induction

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## Phycomorph Morphogenesis in macroalgae

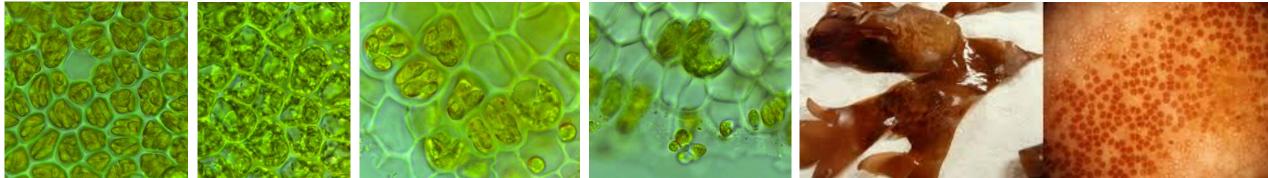


### WG 1: Fertility induction



- Why is fertility induction important for aquaculture?
- How has Phycomorph advanced knowledge of the control of seaweed fertility?
  - Training schools
  - Phycomorph STSM
  - PEGASUS guidelines
  - Scientific progress & papers
  - Dissemination





## Why is this important for aquaculture?

Fertility induction - two 'conflicting' issues:

Controlling lifecycle & the formation of gametes & zygotes for:

- Avoiding biomass breakdown for maximal biomass production (spontaneous sporulations)
- Controlled seeding of cultures



Phycomorph

Fatemeh Ghaderiardakani/AlgaPlus

## WG 1: Training school

- First Workshop & Training School
- Seaweed Cultivation
- 2016
- Kavala, Greece
- Sotiris Orfanitis & Kristos Katsaros
- Avoiding spontaneous sporulation (Ulva)
- Controlling life-cycle and inducing fertility (Kelps)



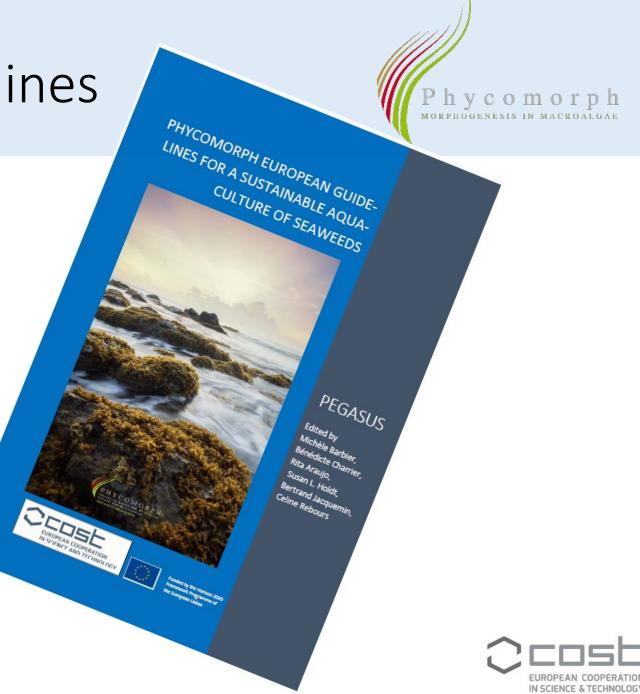
## WG 1: 9 STSMs related to fertility induction

- Neusa Martins: Optimising Kelp Gametophyte culture to investigate transcriptional networks during gametogenesis: CCMAR, PT → AWI, DE
- Patricia Oliviera: Effects of temperature variation on the reproductive stages of *Fucus* sp: Uni Porto, PT
  → Uni Trieste, IT
- Jessica Knoop: Porphyra/cultivation of early life stage: Swansea Uni, UK → Algae+, PT
- Marina Linardic: Induction of fertility of *Dictyota*: Uni Cambridge, UK → Uni Ghent, BE
- Xiaojie Liu: Transcriptomics of Ulva fertility induction: Uni Ghent, BE → Uni Jena, DE
- Yacine Badis: Gamete production *Ectocarpus*: SAMS, UK → Algal Genetics group Roscoff, FR
- Fatemeh Ghaderiardakani: Ulva production: Uni Birmingham, UK → Algae+, PT
- Daniele Grech: Methyl Jasmonate-mediated conceptacle development in red corraline algae: Uni Trieste, IT → Uni Las Palmas, ES
- Omri Nahor: Starch production during gametogenesis in Ulva: Uni Tel Aviv, IL → Uni Jena, DE

## WG 1: PEGASUS guidelines

Contributions from Phycomorphers from science and industry

- also on fertility induction



## WG 1: PEGASUS guidelines

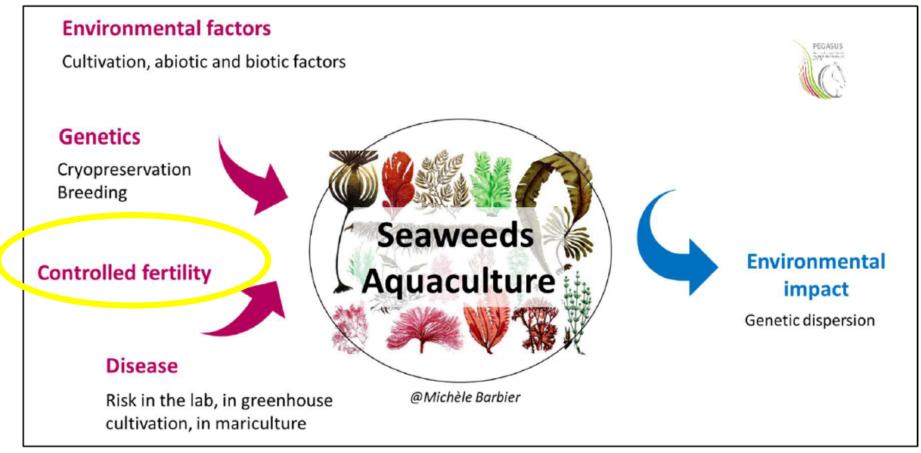
Phycomorph Morphogenesis in Macroalgae

### Controlling fertility = major challenge:

- Cultivation of new species under artificial conditions
- Improve production of juvenile seaweeds

 $\rightarrow$  more R&D

in cooperation between fundamental seaweed biology and aquaculture





### WG 1: Scientific progress & papers

Joint reviews:

**Controls of seaweed reproductive biology** 

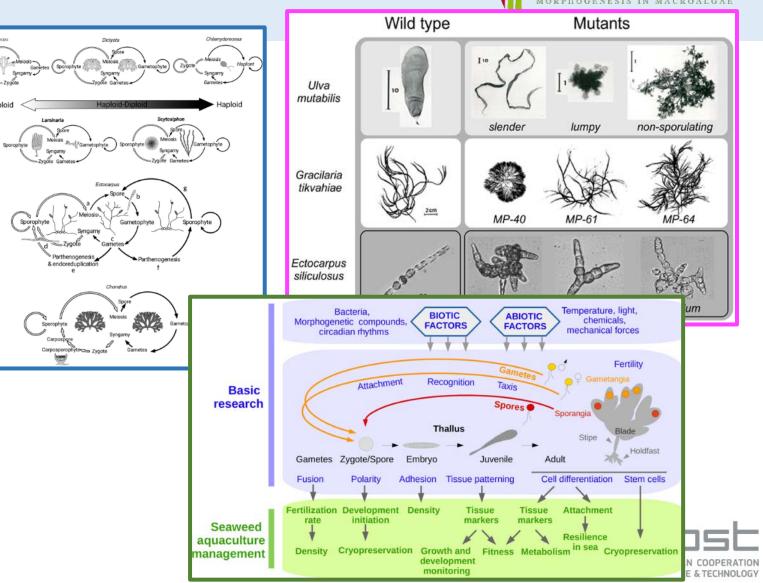
Liu et al. 2017 Botanica Marina

Morphological mutants including non-sporulating mutants

Charrier *et al.* 2017 Frontiers Plant Sci

Fertility bottlenecks (amongst other things)

Charrier *et al.* 2017 New Phytologist



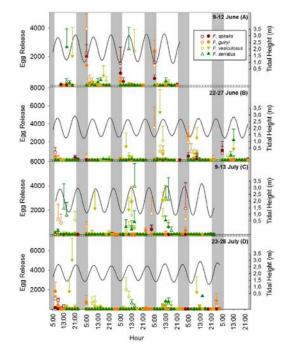
hvcomorph

D1.1: Identify the overall biological and environmental parameters triggering the shift to the reproductive phase (>7 papers)

#### Pearson Group: brown algae

Timing of gamete (egg) release in co-existing *Fucus* species (in natural populations) leads to partial reproductive barriers between species.

#### Monteiro et al. Scientific Reports

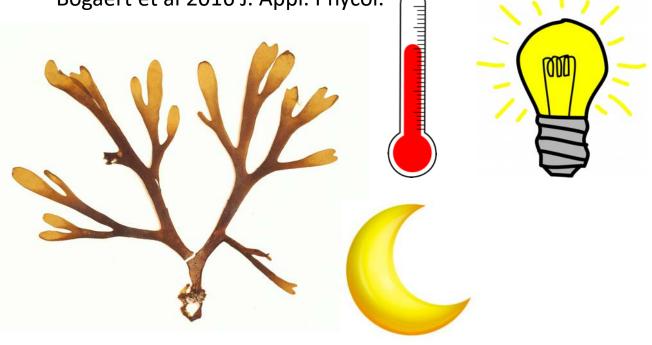


### De Clerck group: brown algae

Regulation of gamete- and spore release by abiotic factors in *Dictyota* 

hycomorph

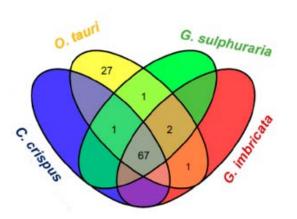
Bogaert et al 2016 J. Appl. Phycol.



D1.2: Identify the chemical compounds and signalling molecules (morphogens) mediating the differentiation of the morph reproductive cells (gametes or spores) (>5 papers)

### Robaina Group: Grateloupia

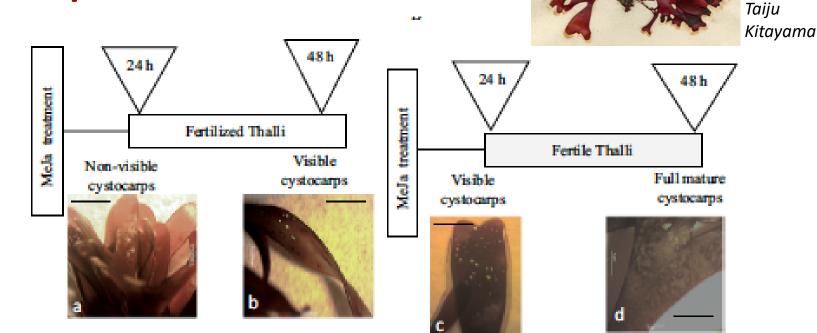
A) METABOLIC PATHWAYS



**Transcriptome:** Polyamines, ethylene, methyl jasmonate.

New ways to enhance reproduction.

Garcia-Jiminez et al 2018, Marine Drugs.

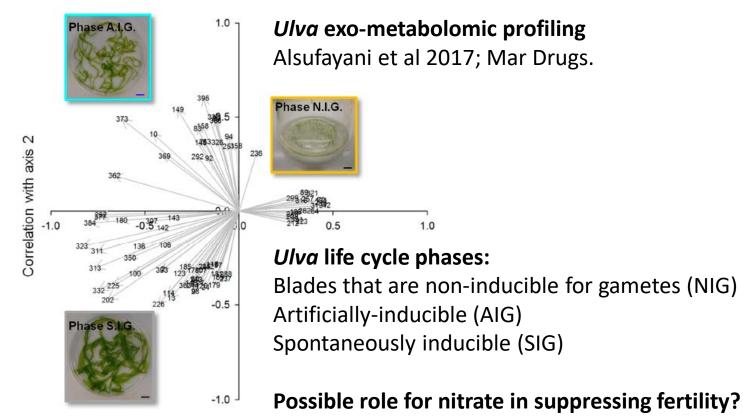


Methyl Jasmonate: Garcia-Jiminez et al 2017, J Phycol.

Ethylene-induced genes: Garcia-Jiminez et al 2018, J Phycol.

D1.3: Identify the mode of action of the signalling molecules within the whole seaweed tissue: localisation of biosynthesis, morp transport and receptors (>2 papers)

### Wichard Group: Ulva



Correlation with axis 1

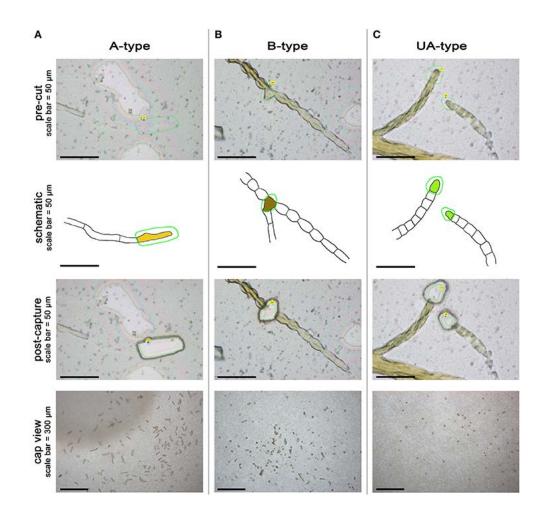
D1.4: Characterize the cell differentiation steps leading to the development of reproductive organs, at both the cellular (microscopy) and the transcriptional level (microdissection followed by Phycomorphics) (> 2 papers)

### **Charrier group: brown algae**

First step – a PHYCOMORPH tool that can be applied to seaweed fertility?

Laser capture microdissection in *Ectocarpus siliculosus*: the pathway to cellspecific transcriptomics in brown algae.

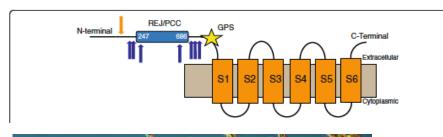
Saint-Marcoux et al. 2015 Frontiers Plant Sci



D1.5. Identify cell markers (transcripts and proteins) allowing early identification of reproductive cells prior to morphological differentiation (>2 papers)

### De Clerck group: brown algae

*Ectocarpus* male-specific gamete genes Lipinska et al 2016; BMC Evol Biol

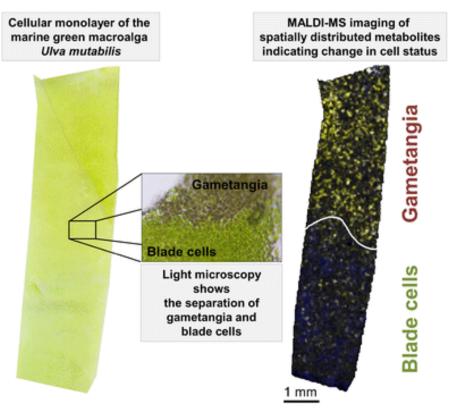




### Wichard Group: Ulva

Gametogenesis biomarkers identified:

Kessler et al 2017; Anal. Bioanal. Chem.



## WG 1: Dissemination & outreach

Example:

4<sup>th</sup> WG meeting in Grenaa Denmark in connection with:

- 8<sup>th</sup> Nordic Seaweed Conference "From research to industry"
- Regional industry network meeting

- Site visit at commercial IMTA site
- Participants from DK/EU industry at WG meeting
- PEGASUS promoted at conference
- Phycomorph MCs and IP = speakers at conference
- Two new spin-off companies benefitting from Phycomorph network







## WG 1 Speakers: Early Carrier Investigators of the second o

- 1. Xiaojie Lui (Ghent University, Belgium): Gene expression analysis of gametogenesis in *Ulva mutabilis*
- 2. Peter Søndergaard Schmedes (DTU Aqua, Denmark): Fertility and the induction of tetraspores in *Palmaria palmata*

